ROXBURGHINOL, A 1,2-DIHYDROANTHRAQUINONE FROM THE LEAVES OF CASSIA ROXBURGHII

D. ASHOK and P. N. SARMA

Department of Chemistry, Osmania University, Hyderabad-500 007, India

(Revised received 18 April 1985)

Key Word Index—Cassia roxburghii; Leguminosae; leaves; 1,2-dihydroanthraquinone; 1,2-dihydro-1,3,8-trihydroxy-2-methyl anthraquinone; roxburghinol.

Abstract—A new 1,2-dihydroanthraquinone, roxburghinol, was isolated from the leaves of *Cassia roxburghii*. Its structure was established as 1,2-dihydro-1,3,8-trihydroxy-2-methyl anthraquinone on the basis of physical and chemical evidence.

INTRODUCTION

The genus Cassia elaborates several types of hydroxyanthraquinones [1]. In the present communication the isolation and structural elucidation of a new anthraquinone designated as roxburghinol (1) from the leaves of C. roxburghii Benth, is reported.

RESULTS AND DISCUSSION

Roxburghinol (1), mp 211°, $[\alpha]_D^{25}$ – 12.2°, $C_{15}H_{12}O_5$, $[M]^+$ 272, gave a pink colour with alcoholic magnesium acetate indicating it to be a hydroxyanthraquinone [2]. It exhibited reversible oxidation-reduction with aqueous sodium dithionite and air [8], and also gave an intense red colour with aqueous sodium hydroxide indicating the presence of a hydroxyquinone system [3]. Its phenolic nature was indicated by its solubility in aqueous alkali and red colour with alcoholic FeCl₃. It formed a triacetate (2), mp 170°, C₂₁H₁₈O₈, [¹H NMR (90 MHz, CDCl₃); three OAc at δ 2.42, 6H, s and 2.15, 3H, s] on treatment with acetic anhydride and perchloric acid indicating the presence of one alcoholic and two phenolic hydroxyl groups. Its UV spectrum, $\lambda \underset{\text{max}}{\text{Dioxane}}$ nm (log ε): 240 (4.16), 245 (4.15), 252 (4.13), 258 (4.18), 262 (4.16), 287 (3.9), 424 (4.0), was similar to those of 1,8-dihydroxy-2-methylanthraquinones [4]. In the IR spectrum (KBr) of 1, absorptions due to phenolic hydroxyl (3320 cm⁻¹), chelated carbonyl (1625 cm⁻¹) and non-chelated carbonyl (1672 cm⁻¹) were readily discernible. The appearance of $[M-2]^+$ (100%)

and $[M-H_2O]^+$ (5%) peaks in the mass spectrum of 1 supported a dihydroanthraquinone moiety in 1. The foregoing indicated that roxburghinol was a trihydroxy-dihydroanthraquinone containing a chelated and non-chelated carbonyl. The compound in ethanolic solution formed a complex with cupric sulphate showing the presence of at least one hydroxyl group at an α -position [5]. The presence of a β -hydroxyl was indicated by the solubility of the compound in 5% aq. sodium carbonate solution [3, 6].

The ¹H NMR (300 MHz, CDCl₃) spectrum of roxburghinol (1) contained five signals in the aromatic region. A singlet at δ 7.23 (1H) was assignable to the C-4 proton of the anthraquinone [7]. The spectrum also revealed the presence of two doublets, one at δ 7.76 (1H, d, J=8 Hz) and the other at 7.33 (1H, d, J=8 Hz) assignable to H-5 and H-7 respectively [7]. Further a triplet at δ 7.65 (1H, t, J=8 Hz) was assignable to H-6 [7]. Signals at δ 7.52 (1H, s, D₂O exchangeable) and 11.8 (1H, s, D₂O exchangeable) were assignable to C-3 and C-8 hydroxyls respectively.

In the aliphatic region of the ¹H NMR spectrum of naturally occurring anthraquinones, usually only one signal is observed between $\delta 2.3$ and 2.5 due to aromatic =C-Me. The multiplicity of the signals in the aliphatic region of the NMR spectrum of 1 revealed the presence of

a –CH(OH)–CH–Me moiety. The protons of the methyl group resonated as a doublet at $\delta 1.18$ (J=8 Hz). A broad doublet which appeared at $\delta 5.3$ (1H, J=15 Hz) was

I R=H

2 R = Ac

3

attributed to the hydrogen of the C-1 hydroxyl group. Further a multiplet at δ 2.5 and an ill defined doublet at 4.58 integrating for one proton each were assigned to H-2 and H-1 respectively.

In the ¹H NMR spectrum (90 MHz, CDCl₃) of acetate 2 the following signals were observed: δ 1.3 (3H, d, J = 8 Hz, Me), 1.64 (1H, m, H-2), 2.15 (3H, s, C-1 OAc), 2.42 (6H, s, C-3 and C-8 OAc), 5.18 (1H, ill defined doublet, H-1), 7.32 (1H, s, H-4), 7.34 (1H, dd, J = 8, 2.5 Hz, H-7), 7.72 (1H, t, J = 8 Hz, H-6), 8.16 (1H, dd, J = 8, 2.2 Hz, H-5). The foregoing data permitted two alternative isomeric structures (1 and 4) for roxburghinol.

DDQ oxidation of 4 would yield 1,3,5-trihydroxy-2methylanthraquinone mp 280° (lit. [9]), whereas 1 would 1,3,8-trihydroxy-2-methylanthraquinone 230-232° (lit. [10, 11]). DDQ oxidation of roxburghinol (1) in benzene afforded 1,3,8-trihydroxy-2-methylanthraquinone (3), mp 229°, $C_{15}H_{10}O_5$, $[M]^+$ 270. The IR spectrum (KBr) exhibited absorptions due to phenolic hydroxyls (3350 cm⁻¹), and chelated (1625 cm⁻¹) and non-chelated (1675 cm⁻¹) carbonyl groups. Its UV spectrum, \(\lambda_{\text{max}}^{\text{MeOH}} \text{nm: 224, 252, 285 and 427, was in close} \) agreement with that reported for 1,3,8-trihydroxy-2methylanthraquinone ($\lambda_{\text{max}}^{\text{MeOH}}$ nm: 220, 255, 285 and 427) [10, 11], as were the ¹H NMR data and the mass spectral fragmentation pattern [10] (see Experimental). However, a direct comparison could not be made for want of an authentic sample of 1,3,8-trihydroxy-2-methylanthraquinone. On the basis of these findings the 1,2-dihydro-1,3,8-trihydroxy-2-methylanthraquinone structure was assigned to roxburghinol and the other isomeric structure (4) was discarded. The mass spectral fragmentation of roxburghinol (see Experimental) was in good agreement with the assigned structure (1). The m/z peak at 270 (100%) arising by loss of two mass units from the parent ion (272) is of diagnostic value in distinguishing 1,2-dihydroanthraquinones from the corresponding fully aromatic anthraquinones. The 13C NMR spectrum of 1 showed the appropriate absorbances for 15 chemically different carbon atoms, the chemical shifts of which were in accordance with the proposed structure (1). The signals at δ 62.25 (C-1), 32.25 (C-2), 18.67 (Me) provided further support for a -CH(OH)-CH-Me moiety in 1. The stereochemistry at C-1 and C-2 could not be settled, since the spectra of reference compounds are not available.

This is the first report of the isolation and characterization of a 1,2-dihydroanthraquinone from natural sources.

EXPERIMENTAL

The air dried leaves (1.5 kg) of *C. roxburghii*, collected locally, were coarsely powdered and extracted successively with petrol (60-80°), CHCl₃ and Me₂CO in a Soxhlet extractor until the

extracts were almost colourless. The petrol extract was concd under red. pres. to a small vol. (250 ml) and left overnight. The waxy material (50 g) which separated was filtered off and the filtrate further concd under red. pres. to furnish a green semi-solid (5 g) which was subjected to CC on silica gel (200 mesh). Fractions of 100 ml each were collected. The column was successively eluted with petrol (fractions 1-8), C₆H₆ (fractions 9-18), C₆H₆-CHCl₃ (1:1) (fractions 19-24), C₆H₆-CHCl₃ (2:8) (fractions 25-29), CHCl₃ (fractions 30-43), CHCl₃-EtOAc (8:2) (fractions 44-53), CHCl₃-EtOAc (1:1) (fractions 54-87), EtOAc (fractions 88-124), EtOAc-Me₂CO (1:1) (fractions 125-138), and Me₂CO (fractions 139-160). Each fraction was monitored by TLC and similar fractions were combined. Fractions 1-8 yielded aliphatic esters (1.2 g). Fractions 9-42 on rechromatography over silica gel (200 mesh) gave β -sitosterol (1.5 g) (mp 137°, $[\alpha]_D$ -36.0° , lit. [12], mp 139° [α]_D -36.0°). Fractions 44–53 gave an anthraquinone-containing orange semi-solid material (250 mg). Fractions 54-124 yielded a mixture of quinonoid compounds (450 mg) whose separation is in progress. Fractions 125–160 yielded a red resinous material (1.1 g).

The anthraquinone-containing material (250 mg) was rechromatographed over silica gel (200 mesh). Fractions of 100 ml each were collected. The column was eluted with C₆H₆ (fractions 1-3), C₆H₆-EtOAc (1:1) (fractions 4-12) and EtOAc (fractions 13-20). Fractions 1-3 yielded negligible amounts of material and were not further investigated. Fractions 4-12 were combined, on the basis of identical TLC behaviour, and on concn furnished an orange solid (205 mg) which was crystallized from CHCl₃ to yield roxburghinol as orange crystals (200 mg), mp 211°, $[\alpha]_D^{25}$ – 12.2° (MeOH; c 0.54) (Found: C, 66.15; H, 4.42. C₁₅H₁₂O₅ requires: C, 66.17; H, 4.41 %); EIMS (probe) 70 eV, m/z (rel. int.): 272 [M] (25), 270 $[M-2H]^+$ (100), 254 $[M-H_2O]^+$ (5), 242 $[270-CO]^+$ (20), 241 $[270-CHO]^+$ (94), 214 $[242-CO]^+$ (8), 213 $[241-CO]^+$ (16), 186 $[214-CO]^+$ (8), 185 $[213-CO]^+$ (8), 158 [186 - CO]+ (5), 157 [185 - CO]+ (8), 129 [158 - CHO]+ (10), $128 [157 - CHO]^+$ (12), $127 [128 - H]^+$ (23), 121 [242/2]doubly charged] + (25), 114 [129 - Me] + (6), 107 [214/2 doubly charged]⁺ (6), 63 $[114 - C_4H_3]^+$ (18); ¹³C NMR (100 MHz, DMSO-d₆): δ191.52 (C-9), 181.24 (C-10), 153.71 (C-8), 152.7 (C-3), 137.27 (C-6), 133.24 (C-5a), 133.14 (C-4a), 124.35 (C-7), 120.78 (C-4), 119.37 (C-5), 116.67 (8a), 114.27 (C-1a), 62.25 (C-1), 32.25 (C-2), 18.67 (Me). The CHCl₃ extract (5.5 g) when chromatographed over silica gel (200 mesh) also gave roxburghinol (150 mg). The Me₂CO extract yielded a water-soluble phenolic compound whose structure elucidation is in progress.

Roxburghinol triacetate (2). To roxburghinol (40 mg) dissolved in 1 ml of Ac_2O , 4–5 drops of $HClO_4$ was added, and the mixture left at room temp. for 48 hr. The triacetate was purified by prep. TLC (silica gel G), using C_6H_6 -EtOAc (9:1). The band at R_f 0.34 was located under UV, scraped off and extracted with CHCl₃. The product crystallized from C_6H_6 as yellow crystals (35 mg), mp 170°. (Found: C, 63.33; H, 4.53; $C_{21}H_{18}O_8$ requires C, 63.31; H, 4.52%) MS m/z (rel. int.): 398 [M]⁺ (<1) 356 [M - CH₂=C=O]⁺ (8), 354 [356 - 2H]⁺ (31), 312 [354 - CH₂=C=O]⁺ (31), 270 [312 - CH₂=C=O]⁺ (100), 242 [270 - CO]⁺ (6), 241 [270 - CHO]⁺ (18).

DDQ oxidation of roxburghinol (1). Roxburghinol (40 mg) in dry C_6H_6 (100 ml) and DDQ (0.1 g) was heated on a water bath for 4 hr. The clear soln was subjected to CC on silica gel (200 mesh). Fractions 25 ml each were collected. The column eluted with C_6H_6 (fractions 1–5), CHCl₃-EtOAc (1:1) (fractions 6–15), EtOAc (fractions 16–25), and Me_2CO (fractions 26–30). Fractions 16–25 were combined, on the basis of identical TLC behaviour, and on concn furnished an orange solid, which was crystallized from Me_2CO to give 1,3,8-trihydroxy-2-methyl anthraquinone (3) as orange crystals, mp 229°, lit. [10, 11], mp

230–232°. (Found: C, 66.67; H, 3.68. $C_{15}H_{10}O_5$ requires C, 66.66; H, 3.70%.) [M] + 270. IR ν_{max}^{KBr} cm⁻¹: 3350, 1675, 1625, 1575, 1570, 1545, 1460, 1420, 1390, 1295, 1280, 1210, 1175, 1163, 1095, 1065, 1045, 1020, 915, 875, 842, 829 and 760; ¹H NMR (90 MHz; DMSO- d_6): δ 6.9–7.2 (2H, H-4 and H-7), 7.3–7.6 (2H, H-5 and H-6) and 2.3 (3H, s, Me); EIMS (probe) 70 eV, m/z (rel. int.): 270 [M] + (100), 242 [M - CO] + (24), 241 [M - CHO] + (95), 214 [242 - CO] + (10), 213 [241 - CO] + (16), 186 [214 - CO] + (5), 185 [213 - CO] + (8), 158 [186 - CO] + (5), 157 [185 - CO] + (9), 129 [158 - CHO] + (10), 128 [157 - CHO] + (12), 127 [128 - H] + (26), 121 (242/2 doubly charged] + (25), 114 [129 - Me] + (7), 107 [214/2 doubly charged] + (10), 63 [114 - C₄H₃] + (15).

Acknowledgements—We are grateful to Professor M. L. N. Reddy, Head of the Department of Chemistry, for providing facilities, Dr. R. S. Kapil, Central Drug Research Institute, Lucknow, for ¹H NMR spectra and Professor Goverdhan Mehta, University of Hyderabad, for ¹³C NMR spectra. We are also grateful to Dr. M. Prabhakar, Department of Botany, Osmania University for identifying and providing the plant material. One of us (D.A.) is grateful to the C.S.I.R., New Delhi, for the award of a Junior Research Fellowship.

REFERENCES

- Thomson, R. H. (1971) Naturally Occurring Quinones, p. 367. Academic Press, London.
- Shibata, S., Takito, M. and Tanaka, O. (1950) J. Am. Chem. Soc. 72, 2789.
- Thomson, R. H. (1971) Naturally Occurring Quinones, p. 40. Academic Press, London.
- Thomson, R. H. (1971) Naturally Occurring Quinones, p. 59.
 Academic Press, London.
- 5. Somogyi, (1952) J. Biol. Chem. 19, 195.
- 6. Graebe, (1906) Annalen 211, 349.
- Thomson, R. H. (1971) Naturally Occurring Quinones, p. 74. Academic Press, London.
- Feigl, F. and Anger, V. (1966) Spot Tests in Organic Analysis, p. 336. Elsevier, Amsterdam.
- Yoshio, H., Naohisa, S., Eiko, K. and Susumu, N. (1962) Chem. Pharm. Bull. (Tokyo) 10, 634.
- Kaur Duggal, J., Yadava, V. S. and Misra, K. (1982) Proc. Nat. Acad. Sci. India. 52A, 189.
- Mulchandani, N. B. and Hassarajani, S. A. (1977) Planta Med. 32, 357.
- Devon, T. K. and Scott, A. I. (1972) Hand Book of Naturally Occurring Compounds, Vol. II, p. 305. Academic Press, New York